# HOW TO PRESENT AT A SCIENTIFIC MEETING

Dr. Fern Tsien Department of Genetics LSUHSC

# Important Deadline #1:

- Abstracts are due on or before Friday, July 18<sup>th</sup> by 5:00!!!
- Medical student abstracts are due on August 1<sup>st</sup>.
- Follow the guidelines sent to you.
- DO NOT change the font size or style.
- We will use the abstract you send us to generate the Abstract Book to give out during the poster session and to the judges ahead of time.

# What is an Abstract?

- An abstract is a one-page summary of your project.
- List your name, mentor's name, etc. as described in the template.
- Affiliations: department and school

- Use only the template we provide.
- <u>This template has the correct sized fonts and sizes we will use.</u>
   <u>Do not change the font or size!</u>
- Make sure your mentor approves of your abstract before you send it to us!
- When you submit your abstract, please be sure to save the file with your last name listed first. For example: ShieldsHeatherAbstract.docx
- Send it to: Wanda Joseph <u>wjose3@lsuhsc.edu</u>
   <u>AND</u> Brianne Jones <u>bjon13@lsuhsc.edu</u>

# Your Name (first, middle initial, last)

Classification (High School, Undergraduate, Medical) Name of School, City, State

Mentor's Name: Mentor's Affiliation (LSUHSC, Tulane SOM, Xavier, Children's Hospital, etc.)

## "Title of Project"

Abstract (summary of project, not to exceed one page)

Body of Abstract: Left Justified, 11 pt Arial font.

#### Charity F. Sylvester Undergraduate Xavier University, New Orleans, LA

Mentor: Imran Mungrue, Ph.D. Louisiana State University Health Sciences Center, Department of Pharmacology and Experimental Therapeutics

#### "Assessing SNPs in the ABCC6 Transporter in an Acadian Family Predisposed to Cardiovascular Disease"

Cardiovascular disease (CVD) encompasses pathologies of the cardiovascular system, which includes diseases of the heart and arteries. Many factors, including genetics, behavior, ethnicity and environment are known to contribute to the disease progression. However, only about 10% of the genetic causes of CVD have been defined. Pseudoxanthoma elasticum (PXE) is a genetic disorder that causes calcified skin lesions known as pseudoxanthomas, retinal deterioration, or expedited arteriosclerosis. PXE has been linked to a mutation in ABCC6, which has also been associated with an increase in coronary artery disease. We found an Acadian family predisposed to premature cardiovascular disease, with eight family members experiencing my ocardial infarction prior to age 43. A genetic cause for this familial aggregation is not yet known. We hypothesized that a SNP in the gene ABCC6 (ATP Binding Cassette Subfamily C Member 6) could contribute. The function of ABCC6 is currently unknown.

In our study, we sought to determine whether members of the family possessed a SNP, (rs726537060), which results in a nonsense mutation in ABCC6 in which arginine is substituted for a termination amino acid at codon 1141. The alleles associated with this SNP are cytosine (C) and thymine (T). Cytosine is the major allele and thymine is the pathogenic, minor allele. Thymine has a 3% minor allele frequency. People who are affected by PXE possess a homozygous recessive genotype at the SNP, but studies suggest that a heterozygous genotype can cause symptoms associated with PXE such as premature atherosclerosis.

We found in all the samples studied that they did not posses the pathogenic allele. This means none of the family members studied expressed the pathogenic allele. We conclude that the R1141X SNP in the ABCC6 gene is not a genetic factor causing premature cardiovascular disease in the Acadian family. Further studies will focus on global SNP associations.



#### Supernumerary marker chromosome (SMC) 17: new case report, delineation of the phenotype, and comparison with other segmental 17p duplications.

Trisomy of the short arm (p arm) of chromosome 17 resulting from a supernumerary marker chromosome (SMC) is very rare producing a variety of phenotypes, with some patients often dying at an early age. We present a 3 week old patient with facial anomalies including cleft palate and cardiac defects. High resolution chromosomes and fluorescence in situ hybridization were done which revealed a trisomy of the 17 p arm and part of q arm (the long arm). We intend to compare this new case with other cases of trisomy 17p and display why our patient is unique.

# Important Deadline #2:

Posters are due Wednesday, July 23<sup>rd</sup> at 5:00!

If we do not receive a poster by this date, your mentor will be responsible for the printing of the poster!!!

# Important deadline #3:

# Summer research Internship Poster Day Wednesday, July 30, 2014

 Medical education Building (MEB), 1901 Perdido St, NO, LA 70112

# Schedule:

- 12 noon-1:00 pm, First Floor Lobby Hang up your poster
- 1-2:30 pm, First Floor Lobby Interns and judges only!
- 2:30-3:30, First Floor Lobby Open to the public
- 3:30-4:30 First floor Auditorium A <u>Awards ceremony, open to the public</u>

# Who will be presenting posters?

- All high school and undergraduates in this program are required to present a poster on the 30<sup>th</sup>.
- Since classes begin early for medical students, they will present their posters during the medical research day in the fall.
- Student presentations will be judged and awards will be given for each category

# Preparing the posters

- First and most important: make sure that your mentor approves of the information that will be presented in the poster.
- Second most important: Your name should go first, your mentor's name last, and everyone else who helped you (other students, post-docs, etc.) in the middle. Make sure not to leave out anyone who helped you!
- Make sure that you understand everything you write on the poster. You should be able to explain your project to the judges.
- In general, try to keep text towards the outside and figures and tables in the center.
- The abstract is not necessary for the poster.

# Preparing the posters

- <u>Use the Power Point poster template sent to you by Wanda Joseph or</u> <u>Brianne Jones</u> (not your friends or past interns) with the proper logos.
- These correspond to your mentor's affiliation and the Summer Program funding source.
- The logos on your poster may differ from the ones on your lab mates! Do not change them!
- Use at least a 24 point font size so the text will be visible from 3 feet away.
- Feel free to adjust the box sizes depending on the amount of text or figures you have.
- Use any color you want to. Express yourself! Exceptions:
  - Black or deep blue for background of entire poster.
  - Image enlarged to cover the entire background.
- Spell out any acronyms the first time you use them. People outside of your lab do not know what "DBS" or "FSHD" is.
- Refer to guidelines sent to you.

# Once your poster is done:

• Save it as a PPT or PPTX file.

- When you submit your poster, be sure to save the file with your last name listed first. For example: ShieldsHeatherPoster.pptx
- Send it to: Wanda Joseph wjose3@lsuhsc.edu

# AND Brianne Jones bjon13@lsuhsc.edu

- You will be notified when your poster is ready to pick up from the Genetics office.
- You are responsible for hanging up the poster on July 30<sup>th</sup> at 12:00.
- Plan to take your poster down at the end of the poster session and give it to your mentor. Let us know in advance if you want an extra one for yourschool.
- Posters are due Wednesday, July 23rd!!!
- If we do not receive a poster by this date, your mentor will be responsible for the printing of the poster.

# Next: Practice your presentation

- Practice with your mentor and your lab members!
- Anticipate questions and look up the answers ahead of time
- Practice, practice, practice so you sound polished.
- Practice in front of your friends or in front of a mirror.

# What happens at a poster session?









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# What happens at a poster session?

Please dress appropriately (business attire).

- The posters will be displayed early so the judges will have a chance to see them ahead of time. Also, they will have a copy of the abstracts.
- At 1:00, stand by your poster. Judges will be passing by asking you questions.
- The posters will be judged on poster display and your presentation (enthusiasm, understanding of the topic, etc)
- Practice ahead of time a short (2-5 minute) description of your poster. Sometimes people ask specific questions, sometimes they ask "tell me about your project"
- DO NOT READ THE POSTER TO THE JUDGES
- Think of possible questions you may be asked. If you do not know an answer, it is OK to say "I don't know"
- At 2:30 the posters will be available to the public. Your family is invited.
- At 3:30 we will move to the 1<sup>st</sup> floor auditorium and give out awards!

# Nervousness: How to fight back

- Practice ahead of time. A well organized, practiced talk will almost always go well.
- If you draw a blank, then looking at your poster will help you get back on track.
- Taking a deep breath will calm you down.
- Slow down. Take a few seconds to think about a question that is being asked before you answer it.
- Bring notes. if you are afraid that you will forget a point, write it down on a piece of paper and bring it with you. However, you don't want to have a verbatim copy of your talk, instead write down key phrases that you want to remember to say.
- Be prepared to answer questions. You don't have to know the answer to every question, however you should be prepared to answer questions about your work. Before the poster session, think about what questions you are likely to get, and how you would answer them
- It is okay to say "I don't know" or "I hadn't thought about that, but one possible approach would be to..."

# What is wrong with this poster?



# Effect of Gain-of-Function Mutant Rb on the Sphere-Forming Ability of Cell Lines

Your name goes first, Graduate students and post-docs that helped you, Mentor is last Mentor's department and University



## Abstract

Osteosarcoma, the most common bone cancer, is the second highest cause of cancer-related death in children and adolescents. Approximately 90% of cases show micro-metastasis at diagnosis, making systematic chemotherapy the first choice of treatment. Despite intensive chemotherapy, the survival rate for high-grade osteosarcomas remains at only 50 80%. This persistence is mainly due to the ability of osteosarcoma cells to metastasize and develop resistance to therapy Increasing evidence suggests that cancer stem cells (CSCs) or tumor initiating cells (TICs) are responsible for these properties and that the inadequacy of current treatments may be result from the inability to target CSCs or TICs in eosarcoma. Recently, we have demonstrated that small populations of osteosarcoma cells can grow and form spheres both serum- and anchorage-independent manners. Importantly, as few as 200 cells from these spheres efficiently initiated osteosarcomas in tumor transplantation models. These results suggest that spheres are enriched with osteosarcoma CSCs. Approximately 50% of osteosarcomas contain alterations in the tumor suppressor p53 gene. Many hotspot p53 mutants show oncogenic functions by their gain-of-function phenotypes such as increased transformation, metastasis and drug resistance, which can not be explained simply due to loss of wild-type p53 function. Our preliminary results indicate that downregulation of a gain-of-function mutant p53R172H results in a dramatic reduction of sphere-forming ability of an teosarcoma cell line expressing p53R172H. These results suggest that mutant p53 gain-of-function is involved in sphere-forming ability and possibly CSC-like properties of osteosarcoma. However, the exact molecular mechanisms which contribute to sphere formation and CSC-like properties and the involvement of mutant p53 in these cellular phenotypes remain unclear.

<u>Our lange-erm goal</u> is to identify the molecular mechanism underlying the CSC-like properties of osteosarooma. <u>The</u> <u>adjustment</u> of the minipation of the molecular mechanism underlying the CSC-like properties of esteosarooma. <u>The</u> <u>adjustment</u> of the minipation of the state adjustment of the state adjustment of the state of t

## Introduction

Ostessarcoma is a devasating disease in children and young adults. In approximately '90% of ostessarcoma cases, micrometatases are present during diagnosis, making chemotherapy the first choice of treatment. Despine intensive chemotherapy, the survival rate for high-grade ostessarcomas remnins at only 50-80%. This persistence is musinly due to the adulty of ostessarcoma cells for metatasize and devolper presistance to therapy. Increasing evidence suggests that cancer stem cells (CSCs) or tumor imitating cells (TICs) are responsible for the metastatic and drug-resistant properties of cancer cells and that the inadequery of current treatments for high radio colossoricom any result from the mability to target ostesarcoma CSCs. CSCs represent a small fraction of a trunor's cellular population and have the ability of cancer cells annot solutical in cellular composition to the tumor's cellular population and have the ability of sperante new tumors identical in cellular composition to the tumor of varies. CSCs are presented in the mability to independent, semin-independent cell growth typhere formation; tumor initiation, self-ensevability, and multimage independent, semin-independent cell growth typhere information; the three in the metastation and drug resistant properties. However, the molecular mechanism that regulates CSC-like properties such as high metastatis and drug resistant properties. However, the molecular mechanism that regulates CSC-like properties of ordeoarcoarce metasistant properties.

Cancer can arise through alterations to genes that regulate cell proliferation, apoptosis, and sensescence. The tumore suppressor p53, one of the key guardinan of these events, etern its finations through transactivating numerous downtream targets. Tumor suppressor p53 have a single nucleotide polymorphium (SRP) at colon 72 which is either proline (P) or arginine (R). Recent studies have above that the 72R form is more potent in its ability to induce apoptosis in the D7A building domestication of p53 as a transcription factor, thereby losing its tumor suppressor activity. The importance of p53 mutations is emphasized by the dinard observation that the p53 gene at fact activity. The importance of p53 mutations is emphasized by the dinard observation that the p53 gene is nutled in more than 50% of tumors. Mutations in the p53 gene area also observed in approximately 70% of patients with L-Fraumen syndrom (LES), human familat charce-prote disease. LFS is characterized by cardy onset of various types of tumors, moltang outcoasteroma. Several missense mutations and as R17511, R248W, and R27311, are the hotspot truttons in sporadic cancers and use the grained transformation, metastasia, and drug resistance, which can not be explained simple by loss of via/byte p53 function. The molecular mechanisms underlying the again-of-function patients are interviations are transformed to restrice are also observed 72.8NP affects the mutant p53 sgain-of-function activities remain unclear. Further, although the gain-of-function 72.8NP affects the mutant p53 sgain-of-function of mutant p53 the CS. the contributions of mutant p53 the CS. The CS molecular

## Methods and Materials

Cell lines. Human osteosarcoma cell lines U2OS, SJSA1, Saos-2, MG-63, and KHOS NP were purchased from American Type Culture Collection (ATCC, Manassas, VA).

Sphere enlurne. Cells were counted by tryour-blue staining (Sigma Bitchemicals), and live cells (five per well) were plated on 36%-well thirds-wull attachment plate (Corning Inc., Corning, NY, USA) in sphere-specific media consisting of DMEM F12, progesterence (10 mR), putrescience (50 µR), insolin (12.5 gg ml), transferrin (12.5 gg ml), asdum selenite (12.5 gg ml), Sigma Bischemicals), murne EOF (10 ng ml), and murne bVFF (10 ng ml). Papo Tech, Rocky Hill, NJ, USA). Cells were mantained for 10 – 14 days and fresh aliquots of EOF and bHGF were added three times a week. Sphere formation was observed day using under a phase-contrast microscopy (Nkon Echipor STIO).

Western blotting, MG-63 cells infected with retroviral vectors encoding control empty or a mutan p53 (R175H, E248W, R273H) with a colour 72 polymorphism were based with R174 buffer 50 mM175HC pH 76, 150 mM1 NocL1, mM1 BETDA, 1\*s sodium docycholata, O1+7 rition X-100, 01+8 XD3 supplemented with potease inhibitor cocktail (Roche)(1 mM phenylmethistalforyl fluoristi (PMSF), 02 mM sodium orthorwandste, and 100 mM sodium Ruonin). Whole extracts were spaced by SDE-8740E and transferred on PVDF membranes (GE Healthcore Biosciences). After blocking with 5% non-fat milk in 1 x Tris-buffered siling (TBS) with 01+87 Neeza) (TBSF), block were investued with inhormandia providase (Santa Cruz). To visualize signals, Super Signal West Dura Chemilaminescent subtrates (GPrere Biotechnology) were used according to numaficative instructions. The signals were deteed on ang a Boral Tizzo Loe detection system (Biord)

Sphere culture. Cells were counted by trypan-blue staining (Sigma Biochemicali), and live cells (five per well) were plated on 96-well thralow attachment plate (Corning Inc., Coming, NY, USA) in sphere-specific media consisting of DMEMF12, progesteence (10 nM), putressine (50 M(h), insulin (12 5 gg ml), transferm in (12 5 gg ml), sodum selentic (12 5 ng ml), Sigma Biochemicals), murine EGF (10 ng ml), and nument FGF (10 ng ml), Pepro Tech, Rocky Hill, NJ, USA). Cells were maintained for 10 – 14 days and fresh aliquots of EGF and FFGF were added three time a week. Sphere formation was observed aduly using under a phase-contrast microscopy (Nikon Eclips TS100).

Western bløtting, MG-63 cells infected vith retorvial vectors encoding control empty or a mutan p53 (RJ 75H, R24W, or R227H) with a codon of the control of

Figure 1

#### Western blotting for mutant p53



Figure 2. Western blotting, MG-63 cells were infected with mutant p53-encoding retroviral vectors to generate subcell lines expressing mutant p53. Cells were lysed in RIPA buffer and whole cell extracts were subjected to western blotting for p53 (DOI antiboby) and loading control vinculin. U2OS having wild-type p53 was used as a positive control.

## Results

Our long-term goal is to identify the molecular mechanism underlying the CSC-like properties of osteosarcoma. *The objective of this study*, is to investigate the effects of several hotspot p153 mutants increase the sphere-forming ability of nonano osteosarcoma cell lines. *Que Inpodhesis* is that gain-of-function p53 mutants increase the sphere-forming ability of osteosarcoma cell. To test our hypothesis, we first characterized the sphere-forming ability of several available human osteosarcoma cell lines. Study 155 mutants increase the sphere-forming ability of osteosarcoma cell lines and as U2OS in fly53 wild-type. JISAI (155 wild-type.) Mic fougat that presented of the Sphere-forming ability of osteosarcoma cell lines, site of the sphere-forming ability of several available human osteosarcoma cell lines, sphere formation when 500 cells were tested for 2 weeks of culturing in sphere-specific conditions. These results may suggest that the presence of viid-type p53 is not crucial for the sphere formation. Assays for other cell lines are on-going. We next infected MicG3 ealed line with retroviral vectors encoding p53R175H72P, p53R154W7X1278, p53R1245W7128, p53R2154W71278, p53R2154

# Table 1

#### Table 1. Results of sphere formation assays

Cell lines	p53 status	Cell# examined	# of spheres formed	% sphere formation
U2OS	wild-type	480	0	0
SJSA1	wild-type	480	1	0
Saos-2	null	480	318	66.3
MG63	null	480	0	0
MG63 R175H/72P	R175H/72P	480	84	17.5
MG63 R175H/72R	R175H/72R	480	160	33.3
MG63 R248W/72P	R248W/72P	480	217	45.2
MG63 R248W/72R	R248W/72R	480	144	30.0
MG63 R273H/72P	R273H/72P	480	112	23.3
MG63 R273H/72R	R273H/72R	480	136	28.3
KHOS	R156P	480	112	23.3

# Conclusions

#### Conclusions

- 1. Spheres vary in size and rate of growth in different osteosarcoma cell lines.
- The presence or absence of wild-type p53 does not have any effects on the sphere-forming ability of osteosarcoma cell lines.
- 3. The presence of mutant p53 does enhance the sphere formation of osteosarcoma cells.
- The effects of p53 codon 72 polymorphisms vary in different p53 mutations.
- 5. All p53 mutants confer osteosarcoma cells with sphere-forming abilities.

#### **Future directions**

- 1. Examine the effects of mutant p53 on other CSC-like properties such as tumor initiating ability, self-renewal, metastatic potential, and drug resistance.
- 2. Examine the effects of mutant p53 down-modulation in various osteosarcoma cell lines carrying mutant p53.
- 3. Identify genes that regulate sphere-forming ability and CSC-like properties of osteosarcoma cells.

# Example of a better poster



# **AXIN2** Gene Instability In Colon Cancer

Summer Student (you), People who helped you, mentor Mentor's department and University



## Abstract

Colon cancer is one of the most prevalent and fatal cancers in the world. In the United States, 10% of all cancer patients have colon cancer. The disease begins when adenomatous polyps, fleshy growths that line up on the inside of the colon, become cancerous. Colonoscopy is often performed to detect these polyps. Regular testing after the age of 40 can drastically reduce the risk of developing colon cancer.

The AXIN2 gene, located in the region of 17q23-q25, is a gene of interest due to its interaction with the Adenomatous polyposis coli (APC) gene in the Wnt signaling pathway and its association with colon cancer with defective mismatch repair. Mutations in the Adenomatous polyposis coli (APC) gene have been found in about 85% of colon cancer patients. However, not much is currently know about the role of AXIN2 in colon cancer development. By conducting research on AXIN2, researchers are hoping that this gene may assist in distinguishing different subgroups of colon cancer. For this project, we analyzed two colon cancer cells lines to determine their karyotypic differences and for any 17q23-q25 region abnormalities.

The majority of the metaphase cells from both of the colon cancer cell lines analyzed were aneuploid, with one cell line (SW480) having a dramatically higher number of chromosomes reaching hypertetrapiloid (103 chromosomes). In addition, the SW480 cell line contained some metaphase cells with an extra copy of chromosome 17 with amplification of the 17q23-25 region. This is the gene location of AXIN2, indicating the possibility of AXIN2 over-expression leading to the colon cancer in this cell line.

## Introduction

The colon is the last portion of the large intestine, which also includes the rectum. Colorectal cancer (CRC), also known as colon cancer, is the third most common cancer in the world and the second most fatal cancer in the Western hemisphere. It is reported that approximately 655,000 people worldwide die from this disease every year. It usually arises from adenomatous polyps that line the inside of the colon. Mutations in certain genes are have been associated with this disease.

One significant gene known to cause CRC is the adenomatous polyposis coli gene (APC). The APC gene is located on the chromosome 5 between positions 21 and 22. Its normal function is to provide instructions for the creation of the APC protein, which helps control how and when a cell should divide. Mutations in this tumor suppressor gene can cause CRC, gastric (stomach) cancer, and Turcot syndrome. Approximately 85% of the people who have colon cancer have a mutation in the APC gene. If a person inherits just one defective copy of the gene from one of their parents, then he or she is almost guaranteed that they will develop colon cancer by the age of 40.

A gene that the APC interacts with is the relatively unknown AXIN2 gene, the focus of this project. Located on chromosome 17 between positions 23 and 24, this gene's protein, Axin2, is presumably very important in the regulation of beta-catenin, which is also a function of the APC gene. Since the APC gene and AXIN2 gene interact in the same pathway, it is believed that a mutation to either gene can affect the other gene. About 30% of the people with colon cancer with defective mismatch repair (the mechanism to correct DNA replication errors) have a mutated AXIN2 gene. The region containing the gene shows loss of heterozygosity in breast cancer, neuroblastoma, and other cancers and tumors. Deletions or mutations in this gene can result in truncated proteins which are most likely inactive. There is a possibility that somatic inactivating mutations in AXIN2 can deregulate beta catenin, and therefore, AXIN2 may be tumor suppressor gene.

Colon Cancer Symptoms		
Constipation	<ul> <li>Vomiting</li> </ul>	Stomach cramps
Thin stool	Diarrhea	
Hematochezia (Blood in stool)	Unexplained Weight loss	

### Figure 1

The AXIN2 gene is located on Chromosome 17 on the q arm (long arm) between positions 23 and 24. The gene spans about 35 kbp and 843 amino acids.





Part of a colon with Adenomatous polyps

# Figure 3



Figure 4

#### G-banded Metaphases From Colon Cancer Cell lines



## **Methods and Materials**

Samples and Culture Conditions:

Two colon cancer lines were obtained from human patients. The Sw48 cell line was obtained from an 82 year old female and the SW480 cell line was obtained from a 50 year old male. The cells were grown in DMEM with 10% Fetal Bovine Serum (FBS) and 1% penticillin under normal culturing conditions.

#### **Chromosome Preparation:**

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For solid staining and G-banding, cells were harvested in exponential phase, incubated with colcenid, treated with a KCL hypotonic, and fixed two times with methanol and acetic acid. For solid staining, the cells were dropped onto slides and stained with Giemsa. For G-banding, the cells were dropped noto slides, followed by a short incubation in a trypsin solution prior to staining with Giemsa.

## Results

	Ploidy of Human Colon Cancer Cell Lines				
-		Sw48	Sw480		
table to the right sthe frequency of ent ploidies in the and Sw480 colon ancer cell line.	Total # of cells analyzed	35	20		
	Diploidy = 46 (Normal #) (%)	2 (6%)	0 (0%)		
	Hyperdiploidy 47-57 (%)	33 (94%)	6 (30%)		
	Hypotriploidy 58-68 (%)	0 (0%)	8 (40%)		
	Triploidy = 69 (%)	0 (0%)	0 (0%)		
	Hypertriploidy 70-80 (%)	0 (0%)	1 (5%)		
	Hypotetraploidy 81-91 (%)	0 (0%)	4 (20%)		
	Tetraploidy 92 (%)	0 (0%)	0 (0%)		
	Hypertetraploidy 93-103 (%)	0 (0%)	1 (5%)		
	Sw48 Cell	s	Sw480 Cell		
ded Karyotypes entative of Colon <sup>•</sup> Cell lines. The <b>rows</b> indicate nalities.	28 28- 18 at 28 ann	(14) 57. X-Y, +der 3 -4,+6,+8,+10,	N+ (*) ( N iii 1111 (; iso (1q), +2, iso (3q)) (; iso (1q), +2, iso (3q)) +11,+11,-12, +13, +15		

## **Conclusions and Future Directions**

When compared to normal human diploid cells, the majority of the cells from the Sw48 cell line were hyperdiploids ranging from a total of 47 to 57 chromosomes per cell, while the Sw480 cell line had a wide range of total chromosomes. Journess the start of the sware start is a start of the sware start is a start of the sware start is a start of the sware start of the hypertiplicity, with an extra chromosome 7 in common.

The sv480 cell line was much more unstable in both studies, with common abnormalities including a missing Y, an extra X abnormalX Schremosome, isochromosome ag, and trisony 13, 21, and 22. The previous repert found one extra chromosome 17. However, our results show four 17 chromosomes, with one of them containing additional genetic material at the q23-qter, the critical region of the AXIV2 gene. Fluorescence in *situ* hybridization (FISH), RNA, and protein analyses should be performed to determine the extent of AXIV2 amplification in the Sv480 eell line.

Due to the nature of these immortalized cell lines, shromosome abnormalities are acquired with increased cell proliferation. *In vitro* studies such as this one can help to gives an idea of what can occur *in viro*. More cancer cell lines should be analyzed in order to find genetic differences between the various types of colon cancer.

# Example of a better poster



Expression of *Irf-7* in Plasmacytoid Dendritic Cells is Limited Following Neonatal Respiratory Syncytial Virus Infection

Names Affiliations



Abstract

Nearly all infants are infected with respiratory syncytial virus (RSV) by two years of age. In infants, RSV is the major cause of bronchiolitis and infants who acquire severe RSV bronchiolitis are at risk of developing asthma. Immune protection is incomplete and reinfection is common throughout life. In otherwise healthy adults, RSV infection usually induces wild upper respiratory tract disease. The mechanisms whereby RSV induces severe disease in infants are largely unknown.

We previously found that neonatal, unlike adult, mice fail to induce appropriate antiviral defenses. In particular, type I interferons are not produced in response to RSV infection. As type I interferons are mainly produced by plasmacytoid dendritic cells (pDCs) via interferon regulatory factor 7 (IRF-7: a transcription factor), we hypothesized that neonatal pDCs in response to RSV infection express less *Irf*-7 than adults.

To test this hypothesis, we infected neonatal mice (5d old) and adult mice (7-8wks old) with RSV and purified pDCs from the lung 24h post infection. We then isolated total RNA from the purified pDCs and reverse transcribed the RNA to produce cDNA. Real time PCR was performed with the resulting cDNA to quantify the relative amount of *IJ*-7 in neonatal and adult pDCs.

We found that pulmonary pDCs from naïve neonates express seven fold less  $If_{2}$ ? than pDCs from adults. When infected with RSV, expression of  $If_{2}$ ? in pulmonary pDCs from both neonates and adults increased; however, neonatal pDCs expressed significantly less  $If_{2}$ ? than adults. These data indicate that the muted induction of  $If_{2}^{-2}$  expression in pDCs may play a role in RSV pathogenesis in neonates.

# Introduction



Fig1: RSV induces Type I IFN production in adult pDCs. RSV enters the cells and fuses with endosomal membrane releasing its genomic siRNA. siRNA is recognized by host TLR78 and induces a cascade of signaling events leading to the phosphorylation of IRF-7. Phosphorylated IRF-7 then translocates to the nucleus and promotes the expression of type IIFNs

# Methods



Fig 2: Schematic of the experimental design. Five day old pups or 6-8 wks old adults were infected with RSV or sham infected with media. At one day post infection, total protein was isolated from the lungs of half of the mice. Type I IFNs were measured in the isolated protein using ELISA. The other half of the mice were used for lung pDC purification. RNA were isolated from these purified pDCs and reverse transcribed to cDNA. The resulting cDNA were subjected to real-time PCR to measure the expression of *lif-7* in pDCs.

# Purity of the Isolated pDCs



Fig 3: Purity of the isolated pDCs. Five day old pups and 6-8 wks old adults were infected with RSV. The pDCs were purified using gradient density centrifugation and magnetic head selection. The resulting cells from the purification were then labeled with CD11c and PDCA-1 antibodies to identify pDCs. (A) Purified pDCs from adult lung, (B) Purified pDCs from neonatal lung.

# Neonatal RSV Infection Induced Limited Expression of *Irf-7*



Fig 4: Relative expression of Irf-7 in pulmonary pDCs. Five day old neonates or 6-8 wks old adults were infected with RSV. pDCs were purified at 1 day post infection; and the expression of Irf-7 in these cells were quantified using real time PCR. NS: sham infected neonates; NR: RSV infected neonates; AS: sham infected adults; AR: RSV infected adults.  $^{\circ}$ : p=0.05.

## Neonatal RSV Infection Induced Limited Type I IFNs Response

	IFNα (ng/g lung protein)	IFNβ (ng/g lung protein)
NS	$4.35\pm0.78$	$5.57 \pm 1.13$
AS	3.77± 0.89	$\textbf{8.14} \pm \textbf{2.31}$
NR	5.51± 1.02	$11.8\pm2.43^{\star}$
AR	76.2 ± 11.2*#	$42.3 \pm 5.07 ^{*}\#$

Fig 5: Type 1 IFNs in lung homogenates. Neonatal or adult mice were infected with RSV and total lung protein was isolated using T-Per (Pierce). IFNs and IFNØ were then measured using ELISA at 1 day post-infection. NS: sham infected neonates; NR: RSV infected neonates; AS: sham infected adults; AR: RSV infected adults. \*: p=0.05; NR vs. NS or AR vs. AS; #: AR vs. NR.

# Conclusions

- Neonatal pDCs express less Irf-7 than adult pDCs at baseline.
- RSV infection induces Irf-7 expression in both neonatal and adult pDCs; however, expression of Irf-7 in pDCs from neonates is muted compared to adults.
- $\square$  RSV infection induces limited amount of type I IFNs (IFN $\alpha$  and  $\beta$ ) in neonates.

 The meter muters minera minor of type ITFIS (if its and p) in its mater.
 The meter expression of *Irf-7* and resulting reduction in type I IFNs may play a role in neonatal RSV pathogenesis.

# Acknowledgement

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# Example of a better poster

# RNA Binding ability of FUS mediates toxicity in a *Drosophila* model of ALS

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## Abstract

Amyotophic Lateral Solemaia (ALS) is a lato-enext neurodispensative disorder characterical by the loss of motor neurons. Muldions in Nucleima (NLS) have been identified as a major component in both familal (NLS) and spensitic (SALS) ALS cases. NLS is an NUA-binding protein implicated in several processos like NNA splicing and minerNNA processing. In normal individuals, bit NLS gone is prodominedly localized in all the nucleus; however in ALS patients, NLS becomes reliabilituded to the cytoplasm as vel, which is believed to be a caseswice pathway for ALS.

Ecopie coperation of human AS with ALV-inked mutations in fly eyes assuss moderate be source obtaining or diagnosticities. Here we commission the role of ANA binding is modialing the neurodegenerative effects of mutant PUS via the ANA Accegnition Mobil (RAM). The RAM declares of the domain tool (MAN-D) or by mutation at the discussed by Istail declares of the domain tool (MAN-D) or by mutation at conservation physical within the PUS RAM too leacine (increme as APA). The APA mutations have been previously shown to mitigate XNAb binding ability in a years model of PUS.

We demonstrate that discupsing the KMA-Domain, by way of deletion or by the APA, point multiplex, can suppress the tanisity of PLB. Interesting confects imaging that shown that discupsing the KMA binding-biblity keeps PLB within the nucleosa junities in ALS accord, where NLB is reliablikuted to the syteplasm), further indicating that subsoliular milabeliatation of PLB is a consider pathway for ALS.

In summer, we have identified a means of rescuing phonotype in our Drosophile model of ALS-associated neurodegeneration, which may be relevant for future clinical studies and interventions in ALS.

# Introduction

>Familial (genetic) ALS accounts for ~10% of all ALS cases, with mutations in FUS accounting for ~4-5% of FALS cases.

>Victims of ALS display loss of muscle mass, increased frailty, loss of mobility, and eventually death.

Currently ALS has no definitive treatment in addition to being ultimately fatal, making the study of ALS all the more urgent and important.

>Steve Gleason, former New Orleam Saint and known ALS patient, in a simply a few yeas, has gone from inciting the loudest recorded noise in the Superdome with his blocked punt all the way to a man confined to a wheelchair and deprived of his former stature.

>Knowing that FUS in itself is an RNA-binding protein, we hypothesized that disruption of its RNA binding ability by deletion of the RRM domain or by 4F-L mutations would reduce the taxicity of mutant FUS.

>We started by transfecting neuronal cells with FUS and corresponding FUS mutations. We then tested our hypothesis by creating transgenic lines with a deletion of the RRM domain in FUS entirely (RRM-D). We next narrowed our focus and created transgenic lines in which we mutated 4 conserved phenylalanine residues within the FUS RRM. To leuche (Innown as 4F-L), Both the RRM-D and 4F-L Ines were used in screens in which the FUS trans-gene was expressed in the fly eyes.

# I. FUS Gene Model



Figure 1: In 2005, ALS-causing mutations in the FUS gene were identified and led to a line of thinking that perhaps errors in RNA metabolism, could be involved in ALS pethogenesis.

#### II. A Drosophila model of FUS Lanson N A et al.

Recently, our lab developed a Drosaphila melanogaster (fruit fly) model as a highly useful system for studying FUS-induced proteinopathies such as ALS.

>FIy models of FUS recapitulate several key features of ALS, demonstrating pupal lethality and larval locomotion defects.



III. RNA Binding ability is essential for FUS-related neurodegeneration.

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NewOrleans

Louisiana Vaccine Center





Figure 2: IRM-binding schrift of FUS regulates toxoby and subsatiule localization. (A) Control Imaging: In neuronal cells, VT FUS (3) is produmently nuclear whereas FUS whith ALS-field mutation (3) in extinction and the objective. The FUS was trapped by the motor-worm specific drive (COV)eH, we observed greater labelity enough puper with an ALS-field mutation as opposed to normal excision in VIT or RNA-binding dried as CPUS similarly, we observed that expression of mutater FUS in motor-worm security and the indication of the labelity enough puper with an ALS-field mutation as opposed to normal excision in VIT or RNA-binding dried as compared to normal locations. (C) LIGN through public controls, interesting's RNA-binding incompletel laws also displayed normal locations. (C) LIGN through public drives as displayed normal locations. (C) LIGN through public drives as compared to normal locations. (C) LIGN through public drives as displayed normal locations. (C) LIGN through public to FUS control, PUS and public resource through the RNA domain or by 4-4-finding bioletism REISC or REIC mutations in 0 y eyes led to external eye degreeation. Howards, RNA binding by deleting the REIN domain or by 4-4-finding to resource to reacting as and the reacting as the reactions.)

## Conclusions

>Disrupting the RRMD omain by way of deletion or by 4FL mutations does indeed seem to significantly rescue phenotype in mutated FUS fly eyes.

>F or further research, we want to express RNA-binding deficient FUS mutations in motor neurons of flies and assess neurodegeneration with respect to motility and lavail crawling ability.

>We would also like to further investigate the link between subcellular localization of FUS and its toxi city, a point of interest which showed up in these experiments.

>Lanson, N. A., et al. 2011. A Drosophila model of FUSrelated neurodegeneration reveals genetic interaction between FUS and TDP-43. Human Molecular Genetics 20, 2510-2523.

>Lanson, N.A., Pandey, UB., FUS related proteinopathies: Lessons from animal models, Brain Res. (2012), doi:10.1016.2012.01.039

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Control UAS-FUS UAS-FUS UAS-FUS UAS-FUS WT R521H R521C R518K

Figure 2: Human ALS causing mutations in FUS lead to neurodeceneration in Drosophila. (A) Scenning electron and light micrographs of adult fly eyes in which expression of Wild-Type or mutant FUS is targeted by the eye specific driver GMIR-GAL4: Whereas the eyes of GMR-Gal4 or FUS WT files show proper pigmentation and ommaticial structure, the ever of files econessing mutant FUS show ommaticial degeneration, partial collegee, and loss of eye pigmentation. (B) Contocal Microscopy: Mutated FUS is shown to leak into the cytoplasm whereas WT FUS is shown to be retained in the nucleus.(C) Larval crawling Assay: Ectopic expression of mutant FUS in motor neuronal results in a larval crawing, defect as compared to UAS-FUS WT expressing animals or driver alone control



· Health education with particular emphasis on basic sanitary practices could have far-reaching effects on mainutrition and disease burden throughout the community

symptoms and to determine if implementation of sanitation Infrastructure and education decreases burden of pediatric illness . Further studies could be used to determine the effectiveness of drip infgation programs by monitoring the height and weight of children in participating families





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